

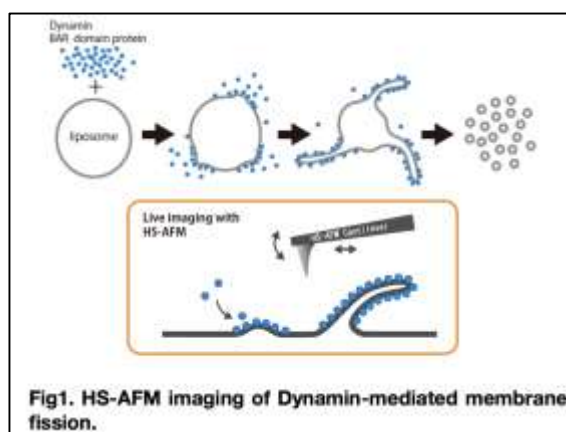
Associate Professor Takeda Tetsuya

Group research activities

Project: Membrane remodeling in health and disease

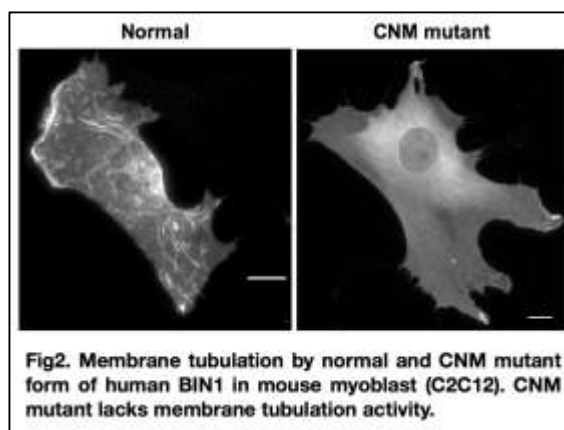
(1) Elucidating membrane fission mechanisms by Dynamin

Endocytosis is required for incorporation of molecules from extracellular environment into cells across membrane barrier. Endocytosis is required for essential biological processes including differentiation, neuronal transduction and signal transduction. In contrast, aberrant endocytosis is tightly linked to various congenital diseases and cancers. Thus, studying endocytosis is important for both physiological and pathological perspectives. In endocytosis, a small area of cell membrane is invaginated and severed to form endocytic vesicles. Dynamin, a large GTPase, plays essential roles in the membrane invagination and fission (membrane remodeling) in endocytosis. Dynamin assembles into an higher ordered “helical” structure at the neck of invaginating endocytic pits and structural changes of the helices coupled with GTP hydrolysis drives membrane fission. However, precise mechanisms of the dyamin-mediated membrane fission remain to be elucidated. We are analyzing mechanism of the dynamin-mediated membrane fission using High Speed Atomic Force Microscopy (HS-AFM) to tackle this fundamental question (Fig.1).



(2) Pathogenesis of congenital myopathy caused by aberrant membrane remodeling

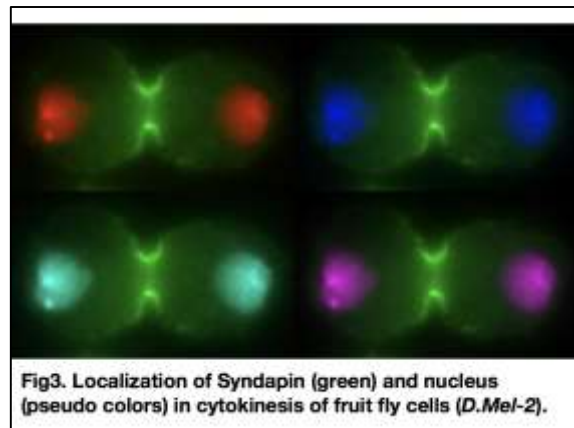
Congenital myopathies are a group of muscle diseases causing muscular weakness. Although frequency of congenital myopathies is relatively low (e.g. 1000 patients in Japan), its pathogenesis is poorly understood thus designated as one of rare intractable diseases by Japanese Ministry of



Health, Labour and Welfare. Centronuclear myopathy (CNM) is one of congenital myopathies and membrane remodeling proteins, Dynamin2 and BIN1, are among causal genes of CNM. We hypothesize that membrane remodeling defects caused by mutations of Dynamin2 and BIN1 leads to pathogenesis of CNM. To elucidate pathogenesis of CNM, we are taking multidisciplinary approaches including cell biology, genetics, biochemistry and biophysics using muscle cell lines and in vitro reconstituted system (Fig.2).

(3) Membrane remodeling in cytokinesis

Cytokinesis is the final step in cell division that physically separates one mother cell into two daughter cells. Regulated cytokinesis is crucial to guarantee proper inheritance of genomic materials into daughter cells. In the development of animals and plants, asymmetrical separation of cell fate determinants is essential for cell differentiation. In contrast, aberrant cytokinesis causes various diseases such as cancer, infertility and developmental defects. Cytokinesis is accompanied by a dynamic morphological changes of cell shape, which is caused by cooperative function of cytoskeleton (actin and microtubules) and membrane. Although function and regulation of cytoskeleton in cytokinesis is relatively well understood, membrane function remains to be elucidated. We have demonstrated membrane remodeling machineries, BAR domain proteins and ESCRT, play essential roles in membrane function in cytokinesis (Takeda *et al.*, *Nat. Cell Biol.* 2004; Takeda *et al.*, *Open Biol.* 2013; Capalbo *et al.*, *Open Biol.* 2012). We are taking multiple approaches including cell biology, in vitro reconstitution and live imaging to elucidate function and regulation of membrane remodeling in cytokinesis.



Selected Publications

- Takeda, T.**, Robinson, I.M., Savoian, M.M., Griffiths, J.R., Whetton, A.D., McMahon, H.T. and Glover, D.M. (2013) *Drosophila* F-BAR protein Syndapin contributes to coupling the plasma membrane and contractile ring in Cytokinesis. *Open Biol.* 7;3(8):130081. (<http://rsob.royalsocietypublishing.org/content/3/8/130081/suppl/DC2>)
- Capalbo, L. Montembault, E., **Takeda, T.**, Bassi, ZI., Glover, D.M., D'Avino, P.P. (2012) The chromosomal passenger complex controls the function of endosomal sorting complex

required for transport-III Snf7 proteins during cytokinesis. *Open Biol.* 2(5):120070.

Takeda, T., Chang, F. (2005) Role of fission yeast myosin I in organization of sterol-rich membrane domains. *Curr Biol.* 15(14):1331-6.

Takeda, T., Kawate, T., Chang, F. (2004) Organization of a sterol-rich membrane domain by Cdc15p during cytokinesis in fission yeast. *Nat Cell Biol.* 6(11):1142-4.

Zhang, Y., Nolan, M., Yamada, H., Watanabe, M., Nasu, Y., *Takei, K. and ***Takeda, T.** (*Corresponding author) Dynamin2 GTPase contributes to invadopodia formation in invasive bladder cancer cells. *Biochem. Biophys. Res. Commun.*, 2016; 480: 409-414.

Other publications

竹田 哲也 (2016) 細胞質分裂における膜ダイナミクスの機能、生物物理 56 (1), 013-017.

Yamada, H., **Takeda, T.**, Michiue, H., Abe, T. and Takei, K. Actin bundling by dynamin 2 and cortactin is implicated in cell migration by stabilizing filopodia in human non-small lung carcinoma cells. *Int J Oncol.*, 2016; 49(3):877-886.

Yamada H, Kobayashi K, Zhang Y, **Takeda T**, Takei K. Expression of a dynamin 2 mutant associated with Charcot-Marie-Tooth disease leads to aberrant actin dynamics and lamellipodia formation. *Neurosci Lett.* 2016 Aug 15;628:179-85.

Yamada, H., Kikuchi, T., Masumoto, T., Fan-Yan Wei, F-Y., Abe, T., **Takeda, T.**, Nishiki, T., Tomizawa, K., Watanabe, M., Matsui, H. and Takei, K. Possible role of cortactin phosphorylation by protein kinase C α in actin-bundle formation at growth cone. *Biol. Cell*, 2015; 107: 1-12.

D'Avino, P.P., **Takeda, T.**, Capalbo, L., Zhang, W., Lilley, K., Laue, E. and Glover, D.M. (2008) Interaction between Anillin and RacGAP50C connects the actomyosin contractile ring with spindle microtubules at the cell division site. *J. Cell Sci.* 121(Pt 8):1151-8.

Glover, D.M., Capalbo, L., D'Avino, P.P., Gatt, M.K., Savoian, M.S. and **Takeda, T.** (2008) Girds 'n' Cleeks o' Cytokinesis: microtubule sticks and contractile hoops in *Drosophila* cell division. *Biochem Soc Trans.* 2008 Jun;36(Pt 3):400-4.

Yonetani, A, Lustig, R.J., Moseley, J., **Takeda, T.**, Goode, B.L. and Chang, F. (2008) Regulation and targeting of the fission yeast formin cdc12p in cytokinesis. *Mol. Biol. Cell* 2008 May;19(5):2208-19.

Noguchi E, Iwama A, Takeda K, **Takeda T**, Kamioka M, Ichikawa K, Akiba T, Arinami T,

Shibasaki M. (2003) The promoter polymorphism in the eosinophil cationic protein gene and its influence on the serum eosinophil cationic protein level. *Am J Respir Crit Care Med.*, 167(2):180-4.

Ueno H, Gonda K, **Takeda, T.**, Numata O. (2003) Identification of elongation factor-1alpha as a Ca²⁺/calmodulin-binding protein in *Tetrahymena* cilia. *Cell Motil Cytoskeleton.* 55(1):51-60.

Takeda, T., Yoshihama, I., Numata, O. (2001) Identification of *Tetrahymena* Hsp60 as a 14-nm Filament Protein/Citrate Synthase-Binding Protein and Its Possible Involvement in the Oral Apparatus Formation. *Genes to Cells* 6 (2), 132-149.

Numata, O., Hanyu, K., **Takeda, T.**, Watanabe, Y. (1998) Tetrahymena calcium-binding proteins, TCBP-23 and TCBP-25. In *Tetrahymena thermophila*, ed. By D.J. Asai and J.D. Forney, *Methods in Cell Biology* 62, 455-465.

Takeda, T., Watanabe, Y., Numata, O. (1997) Direct Demonstration of the Bifunctional Property of *Tetrahymena* 14-nm Filament Protein/Citrate Synthase Following Expression of the Gene in *Escherichia coli*. *Biochem.Biophys.Res.Commun.* 237, 205-210.

Takeda, T., Kurasawa, Y., Watanabe, Y., Numata, O. (1995) Polymerization of Highly Purified *Tetrahymena* 14-nm Filament Protein/Citrate Synthase into Filaments and Its Possible Role in Regulation of Enzymatic Activity. *J.Biochemistry* 117, 869-874.